HCR™ IF protocol for whole-mount zebrafish embryos

This protocol has not been validated for all stages and should only be used as a template.

Technical support
support@molecularinstruments.com

Safety data sheets (SDS)
www.molecularinstruments.com/safety

Patents
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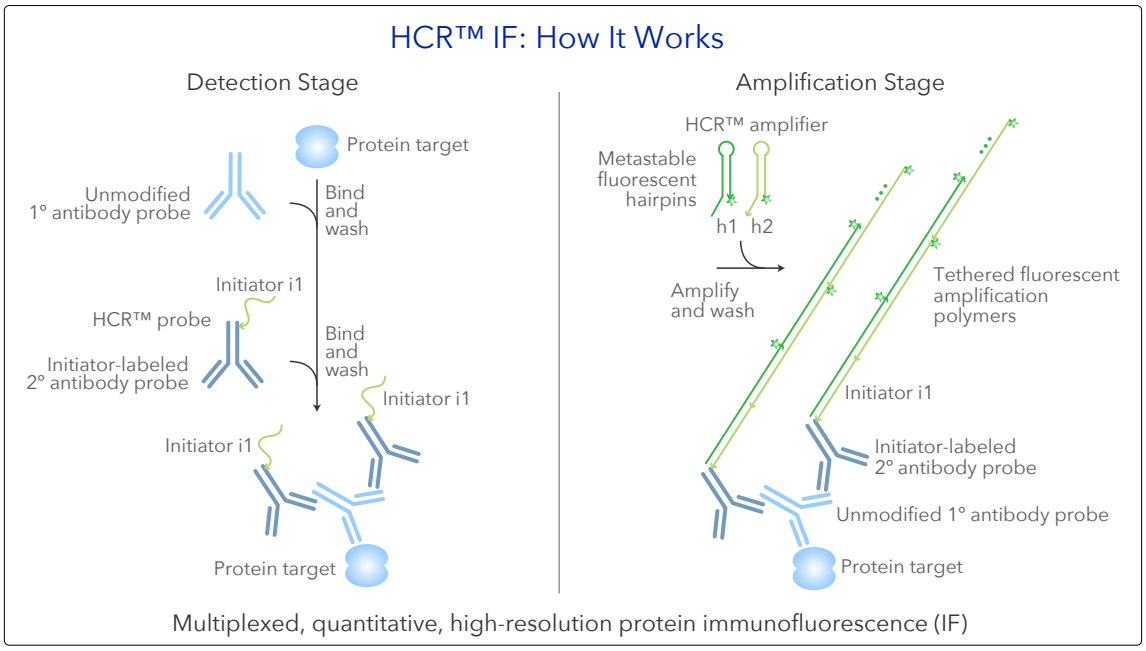

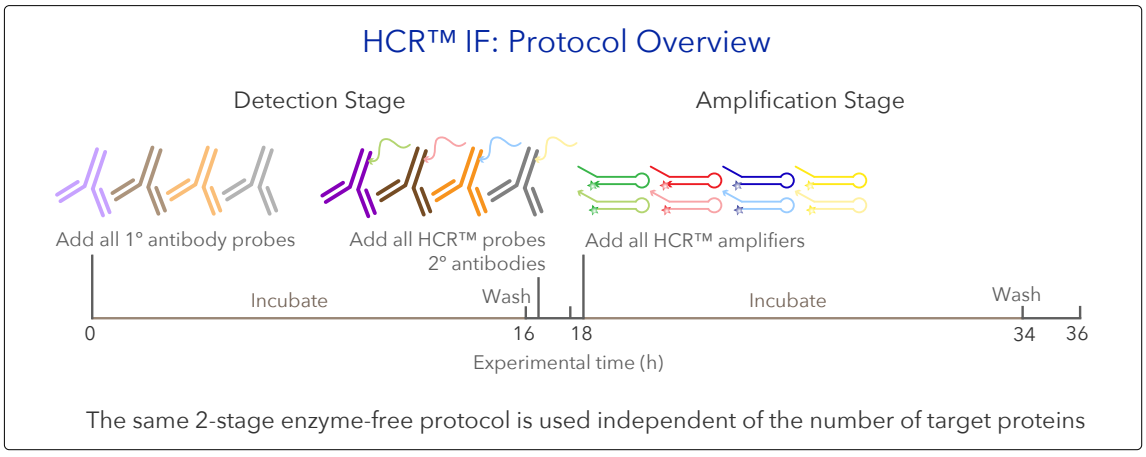

HCR™ IF

Multiplexed, quantitative, high-resolution protein imaging

Multiplexed Experiment

- Order one HCR™ IF bundle per target protein

Example 2-Plex Experiment

- HCR™ IF bundle for target Protein1
  - 1° Ab: Rabbit Anti-Protein1 (your own 1° antibody)
  - HCR™ IF 2° Ab probe: Donkey Anti-Rabbit for use with amplifier B1
  - HCR™ amplifier: B1-647
  - HCR™ IF buffers: antibody buffer, amplification buffer (for use with all bundles)

- HCR™ IF bundle for target Protein2
  - 1° Ab: Mouse Anti-Protein2 (your own 1° antibody)
  - HCR™ IF 2° Ab probe: Donkey Anti-Mouse for use with amplifier B2
  - HCR™ amplifier: B2-488

Storage conditions

- Store HCR™ IF antibody probes and HCR™ amplifiers at -20 °C.
- Store HCR™ IF antibody buffer and HCR™ amplification buffer at 4 °C.
- On the bench top, keep stock solutions on ice.
- Make sure all solutions are well mixed before use.

Preparation of whole-mount zebrafish embryos

1. Collect zebrafish embryos and incubate at 28 °C in a petri dish with egg H_2O.
2. Dechorionate embryos at 27 hpf and wash with fresh egg H_2O.
3. Transfer 40 embryos to a 2 mL eppendorf tube and remove excess egg H_2O.
4. Fix embryos in 2 mL of 4% paraformaldehyde (PFA) for 24 h at 4 °C.
   CAUTION: *use PFA with extreme care as it is a hazardous material.*
   NOTE: *use fresh PFA and cool to 4 °C before use to avoid increased autofluorescence.*
5. Wash embryos 3 × 5 min with 1 mL of 1× phosphate-buffered saline (PBS) to stop the fixation.
   NOTE: *avoid using calcium chloride and magnesium chloride in PBS as this leads to increased autofluorescence in the samples.*
6. Dehydrate and permeabilize with a series of methanol (MeOH) washes at room temperature (1 mL each):
   (a) 100% MeOH for 4 × 10 min
   (b) 100% MeOH for 1 × 50 min
7. Store embryos at -20 °C overnight before use.
   NOTE: *Embryos can be stored for six months at -20 °C.*
8. Rehydrate with a series of graded MeOH/PBST washes for 5 min each at room temperature (1 mL each):
   (a) 75% MeOH / 25% PBST
   (b) 50% MeOH / 50% PBST
   (c) 25% MeOH / 75% PBST
   (d) 5 × 100% PBST

Multiplexed HCR™ IF protocol

Detection stage

1. Block embryos with 500 μL of antibody buffer for 4 h at 4 °C.
2. Transfer 8 embryos to a 1.5 mL Eppendorf tube for each sample.
3. Prepare working concentration of unlabeled primary antibodies in antibody buffer. Prepare 250 μL per sample.
   NOTE: *follow manufacturer's guidelines for primary antibody working concentration.*
4. Remove antibody buffer and add primary antibody solution to embryos.
5. Incubate embryos overnight (>12 h) at 4 °C with gentle rotation (50 RPM).
6. Remove excess antibodies by washing 4 × 30 min with 500 μL of PBT at room temperature.
7. Prepare 1 μg/mL working concentration of initiator-labeled secondary antibodies in antibody buffer. Prepare 250 μL per sample.
   NOTE: *Concentration may be optimized depending on protein target and primary antibody.*
8. Remove PBT and add secondary antibody solution to embryos.
9. Incubate embryos for 3 h at room temperature with gentle rotation (50 RPM).
10. Remove excess antibodies by washing 5 × 5 min with 500 μL of PBT at room temperature.
11. Wash 1 × 5 min with 500 μL of 5× SSCT at room temperature.

Amplification stage

1. Pre-amplify embryos with 350 μL of amplification buffer for 30 min at room temperature.
   NOTE: *equilibrate amplification buffer to room temperature before use.*
2. Separately prepare 30 pmol of hairpin h1 and 30 pmol of hairpin h2 by snap cooling 10 μL of 3 μM stock (heat at 95 °C for 90 seconds and cool to room temperature in a dark drawer for 30 min).
   NOTE: *Hairpins h1 and h2 are provided in hairpin storage buffer ready for snap cooling. h1 and h2 should be snap cooled in separate tubes. This is the amount of hairpins needed for each target in a single sample using 500 μL of incubation volume.*
3. Prepare a 60 nM hairpin solution by adding all snap-cooled h1 hairpins and snap-cooled h2 hairpins to 500 μL of amplification buffer at room temperature per sample.
4. Remove the pre-amplification solution and add the hairpin solution.
5. Incubate the samples overnight (>12 h) in the dark at room temperature.
6. Remove excess hairpins by washing with 500 μL of 5× SSCT at room temperature:
   (a) 2 × 5 min
   (b) 2 × 30 min
   (c) 1 × 5 min
7. Samples can be stored at 4 °C protected from light before microscopy.

Buffer recipes

4% Paraformaldehyde (PFA)	For 25 mL of solution
4% PFA	1 g of PFA powder
1× PBS	25 mL of 1× PBS
	Heat to 50–60 °C to dissolve powder

0.1 M phosphate buffer with Triton X-100 (PBT)	For 40 mL of solution
0.8% Triton X-100	320 μL of Triton X-100
0.061 M K_2HPO_4	0.555 g K_2HPO_4 $3H_2O$
0.039 M KH_2PO_4	0.214 g KH_2PO_4
	Fill up to 40 mL with ultrapure H_2O

5× SSCT	For 40 mL of solution
5× sodium chloride sodium citrate (SSC)	10 mL of 20× SSC
0.1% Tween 20	400 μL of 10% Tween 20
	Fill up to 40 mL with ultrapure H_2O

NOTE: *avoid using calcium chloride and magnesium chloride in PBT as this leads to increased autofluorescence in the samples.*

HCR™ Technology Citation Notes

For citation, please select from the list below as appropriate for your application:

- **HCR™ IF + HCR™ RNA-FISH**
  HCR™ IF + HCR™ RNA-FISH enables a unified approach to multiplexed, quantitative, high-resolution protein immunofluorescence (IF) and RNA fluorescence in situ hybridization (RNA-FISH), with quantitative 1-step enzyme-free signal amplification performed for all protein and RNA targets simultaneously (Schwarzkopf et al., 2021).

- **HCR™ IF**
  HCR™ IF enables multiplexed, quantitative, high-resolution protein immunofluorescence (IF) in highly autofluorescent samples (e.g., FFPE brain tissue sections) (Schwarzkopf et al., 2021).

- **HCR™ RNA-FISH (v3.0)**
  Third-generation HCR™ RNA-FISH (v3.0) enables multiplexed, quantitative, high-resolution RNA fluorescence in situ hybridization (RNA-FISH) with automatic background suppression *throughout the protocol* for dramatically enhanced performance (signal-to-background, subcellular quantitative RNA imaging precision, single-molecule quantitative RNA imaging fidelity) and ease-of-use (no probe set optimization for new targets and organisms) (Choi et al., 2018). Quantitative analysis modes:

  - Subcellular quantitative RNA imaging: analog mRNA relative quantitation with subcellular resolution in the anatomical context of thick autofluorescent samples.
  - Single-molecule quantitative RNA imaging: digital mRNA absolute quantitation with single-molecule resolution in the anatomical context of thick autofluorescent samples.
  - Quantitative RNA flow cytometry: analog mRNA relative quantitation for high-throughput expression profiling of mammalian cells and bacteria.

  Protocols for HCR™ RNA-FISH (v3.0) in diverse organisms are adapted from the Zoo paper.

- **Subcellular quantitative RNA imaging**
  Subcellular quantitative RNA imaging enables mRNA relative quantitation with subcellular resolution in the anatomical context of thick autofluorescent samples (e.g., whole-mount vertebrate embryos). The read-out/read-in analysis framework enables bidirectional quantitative discovery in an anatomical context (Trivedi et al., 2018).

- **Zoo paper**
  Protocols for multiplexed mRNA imaging in diverse sample types (Choi et al., 2016):
  - bacteria in suspension
  - FFPE human tissue sections
  - generic sample in solution
  - generic sample on a slide
  - mammalian cells on a slide
  - mammalian cells in suspension
  - whole-mount chicken embryos
  - whole-mount fruit fly embryos
  - whole-mount mouse embryos
  - whole-mount nematode larvae
  - whole-mount sea urchin embryos
  - whole-mount zebrafish embryos and larvae

- **Single-molecule quantitative RNA imaging**
  Single-molecule quantitative RNA imaging enables RNA absolute quantitation with single-molecule resolution in the anatomical context of thick autofluorescent samples (e.g., 0.5 mm adult mouse brain sections) (Shah et al., 2016).

- **HCR™ northern blots**
  HCR™ northern blots enable multiplexed quantification of RNA target size and abundance for up to 5 target RNAs (Schwarzkopf & Pierce, 2016).

- **HCR™ RNA-FISH (v2.0)**
  Second-generation in situ HCR™ RNA-FISH technology (v2.0) using DNA HCR™ probes and DNA HCR™ amplifiers: 10× increase in signal, 10× reduction in cost, dramatic increase in reagent durability (Choi et al., 2014).

- **HCR™ RNA-FISH (v1.0)**
  First-generation HCR™ RNA-FISH technology (v1.0) using RNA HCR™ probes and RNA HCR™ amplifiers: multiplexed mRNA imaging in whole-mount vertebrate embryos with simultaneous signal amplification for up to 5 target mRNAs (Choi et al., 2010).

- **HCR™ technology**
  HCR™ amplifiers enable multiplexed, quantitative, 1-step, isothermal, enzyme-free signal amplification in diverse technological settings (Dirks & Pierce, 2004).