

HCR™ 2°IF protocol for sample in solution

This protocol has not been validated for all sample types and should only be used as a template.

Technical Support

support@molecularinstruments.com

Safety Data Sheets (SDS)

www.molecularinstruments.com/safety-v3

Patents

www.molecularinstruments.com/patents

Ordering for Multiplex Experiment

Order one HCR™ 2°IF kit per target protein

Example 2-Plex Experiment

- HCR™ 2°IF kit for target Protein1
 - 1° Ab: Rabbit Anti-Protein1 (your own 1° antibody)
 - HCR™ 2° Antibody Probe: Donkey Anti-Rabbit for use with amplifier B1
 - HCR™ Amplifier (v3.0): B1-647
 - HCR™ 2°IF Buffers: HCR™ Antibody Buffer, HCR™ Amplifier Buffer (v3.0) (for use with all kits)
- HCR™ 2°IF kit for target Protein2
 - 1° Ab: Mouse Anti-Protein2 (your own 1° antibody)
 - HCR™ 2° Antibody Probe: Donkey Anti-Mouse for use with amplifier B2
 - HCR™ Amplifier (v3.0): B2-488

Storage conditions

- Store HCR™ 2° Antibody Probes and HCR™ Amplifiers (v3.0) at -20 °C.
- Store HCR™ Antibody Buffer and HCR™ Amplifier Buffer (v3.0) at 4 °C.
- On the bench top, keep stock solutions on ice.
- Make sure all solutions are well mixed before use.

Sample preparation

Samples should be prepared in the same manner as for a traditional immunohistochemistry, up to the primary antibody application step. This may include permeabilization and antigen retrieval. Then proceed with the protocol described below.

Multiplexed HCR™ 2°IF protocol

Detection stage

1. Block samples with 250 μ L of HCR™ Antibody Buffer for 4 h at 4 °C.
2. Prepare working concentration of primary antibodies in HCR™ Antibody Buffer. Prepare 250 μ L per sample.
NOTE: follow manufacturer's guidelines for primary antibody working concentration.
3. Remove HCR™ Antibody Buffer and add primary antibody solution to samples.
4. Incubate samples overnight (>12 h) at 4 °C with gentle rotation.
NOTE: Incubation may be optimized (e.g., 3 h at room temperature) depending on sample type and thickness.
5. Remove excess antibodies by washing 4 \times 30 min with 500 μ L of 1 \times PBST at room temperature.
6. Prepare 1 μ g/mL working concentration of HCR™ 2° Antibody Probes in HCR™ Antibody Buffer. Prepare 250 μ L per sample.
NOTE: Concentration may be optimized depending on protein target and primary antibody.
7. Remove 1 \times PBST and add secondary antibody solution to samples.
8. Incubate samples for 3 h at room temperature with gentle rotation.
9. Remove excess antibodies by washing 5 \times 5 min with 500 μ L of 1 \times PBST at room temperature.
10. Wash 1 \times 5 min with 500 μ L of 5 \times SSCT at room temperature.

Amplification stage

1. Pre-amplify each sample with 250 μ L of HCR™ Amplifier Buffer (v3.0) for 30 min at room temperature.
NOTE: equilibrate HCR™ Amplifier Buffer (v3.0) to room temperature before use.
2. Separately prepare 15 pmol of hairpin h1 and 15 pmol of hairpin h2 by snap cooling 5 μ L of 3 μ M stock (heat at 95 °C for 90 seconds and cool to room temperature in a dark drawer for 30 min).
NOTE: HCR hairpins h1 and h2 are provided in hairpin storage buffer ready for snap cooling. h1 and h2 should be snap cooled in separate tubes. This is the amount of hairpins needed for each target in a single sample using 250 μ L of incubation volume.
3. Prepare a 60 nM hairpin solution by adding all snap-cooled h1 hairpins and snap-cooled h2 hairpins to 250 μ L of HCR™ Amplifier Buffer (v3.0) at room temperature per sample.
4. Remove the pre-amplification solution and add the hairpin solution.
5. Incubate the samples overnight (>12 h) in the dark at room temperature.
6. Remove excess hairpins by washing with 500 μ L of 5 \times SSCT at room temperature:
 - (a) 2 \times 5 min
 - (b) 2 \times 30 min
 - (c) 1 \times 5 min
7. Samples can be stored at 4 °C protected from light before microscopy.

Buffer recipes

1× PBST

1× phosphate buffered solution (PBS)
0.1% Tween 20

For 40 mL of solution

4 mL of 10× PBS
400 μ L of 10% Tween 20
Fill up to 40 mL with ultrapure H₂O

5× SSCT

5× sodium chloride sodium citrate (SSC)
0.1% Tween 20

For 40 mL of solution

10 mL of 20× SSC
400 μ L of 10% Tween 20
Fill up to 40 mL with ultrapure H₂O

NOTE: avoid using calcium chloride and magnesium chloride in PBS as this leads to increased autofluorescence in the samples.

S1 HCR™ Technology Citation Notes

For citation, please select from the list below as appropriate for your application:

- **HCR™ RNA-ISH**

HCR™ RNA in situ hybridization (RNA-ISH) offers unmatched performance, robustness, and versatility imaging RNA targets in diverse organisms and sample types ([Choi et al., 2010](#), [Choi et al., 2014](#), [Choi et al., 2018](#)):

- **HCR™ RNA-FISH**

HCR™ RNA-FISH enables 10-plex, quantitative, high-resolution RNA fluorescence in situ hybridization (RNA-FISH) with automatic background suppression throughout the protocol for dramatically enhanced performance (signal-to-background, quantitative precision, single-molecule fidelity) and ease-of-use (no probe set optimization for new targets and organisms).

- **Enzymatic HCR™ RNA-CISH/RNA-FISH**

Enzymatic HCR™ RNA-ISH integrates enzymatic signal amplification to enable extreme-sensitivity RNA imaging using either chromogenic or fluorescent staining (RNA-CISH/RNA-FISH). In tissue sections, entirely protease-free workflows preserve sample morphology and maintain protein target integrity, enabling seamless compatibility with existing immunohistochemistry (IHC)/immunofluorescence (IF) assays. HCR™ RNA-CISH offers the convenience of brightfield microscopy and the option of archival staining.

- **10-Plex HCR™ Spectral Imaging**

HCR™ RNA-FISH/IF enables quantitative high-resolution imaging of 10 RNA and/or protein targets with 1-step HCR™ signal amplification for all targets simultaneously. The method is suitable even for whole-mounts and delicate samples as it requires no repeated staining, imaging, registration, or stripping ([Schulte et al., 2024](#)).

- **HCR™ RNA-FISH/IF**

HCR™ RNA-FISH/IF enables a unified approach to multiplex, quantitative, high-resolution RNA fluorescence in situ hybridization (RNA-FISH) and protein immunofluorescence (IF), with quantitative 1-step enzyme-free signal amplification performed for all RNA and protein targets simultaneously ([Schwarzkopf et al., 2021](#)).

- **HCR™ IF**

HCR™ IF enables multiplex, quantitative, high-resolution protein immunofluorescence (IF) in highly autofluorescent samples (e.g., FFPE brain tissue sections) ([Schwarzkopf et al., 2021](#)).

- **Subcellular Quantitative RNA and Protein Imaging**

HCR™ RNA-FISH enables analog relative quantitation of RNA and/or protein targets with subcellular resolution in the anatomical context of thick autofluorescent samples (e.g., whole-mount vertebrate embryos) ([Trivedi et al., 2018](#), [Choi et al., 2018](#), [Schwarzkopf et al., 2021](#)).

- **Single-Molecule Quantitative RNA Imaging**

HCR™ RNA-FISH enables digital RNA absolute quantitation with single-molecule resolution in the anatomical context of thick autofluorescent samples (e.g., 0.5 mm adult mouse brain sections) ([Shah et al., 2016](#), [Choi et al., 2018](#)).

- **Read-Out/Read-In Analysis Framework**

The read-out/read-in analysis framework enables bidirectional quantitative discovery in an anatomical context ([Trivedi et al., 2018](#)).

- **Protocols in Diverse Sample Types**

Protocols for HCR™ RNA-FISH and/or IF in diverse sample types are adapted from the *zoo* paper ([Choi et al., 2016](#)):

- bacteria in suspension
- FFPE human tissue sections
- generic sample in solution
- generic sample on a slide
- mammalian cells on a slide
- mammalian cells in suspension
- whole-mount chicken embryos
- whole-mount fruit fly embryos
- whole-mount mouse embryos
- whole-mount nematode larvae
- whole-mount sea urchin embryos
- whole-mount zebrafish embryos and larvae

- **HCR™ RNA Flow Cytometry**

HCR™ RNA Flow Cytometry enables analog RNA relative quantitation for high-throughput expression profiling of mammalian cells and bacteria without the need to engineer reporter lines ([Choi et al., 2018](#)).

- **HCR™ Northern Blots**

HCR™ Northern Blots enable simultaneous quantification of RNA target size and abundance with automatic background suppression throughout the protocol ([Schwarzkopf & Pierce, 2016](#)).

- **HCR™ Amplifiers**

HCR™ Amplifiers enable multiplex, quantitative, 1-step, isothermal, enzyme-free signal amplification in diverse technological settings ([Dirks & Pierce, 2004](#)).