

## HCR RNA-FISH protocol for whole-mount sea urchin embryos (*Strongylocentrotus purpuratus*)

This protocol has not been optimized for all stages and should only be used as a template.

### Technical support

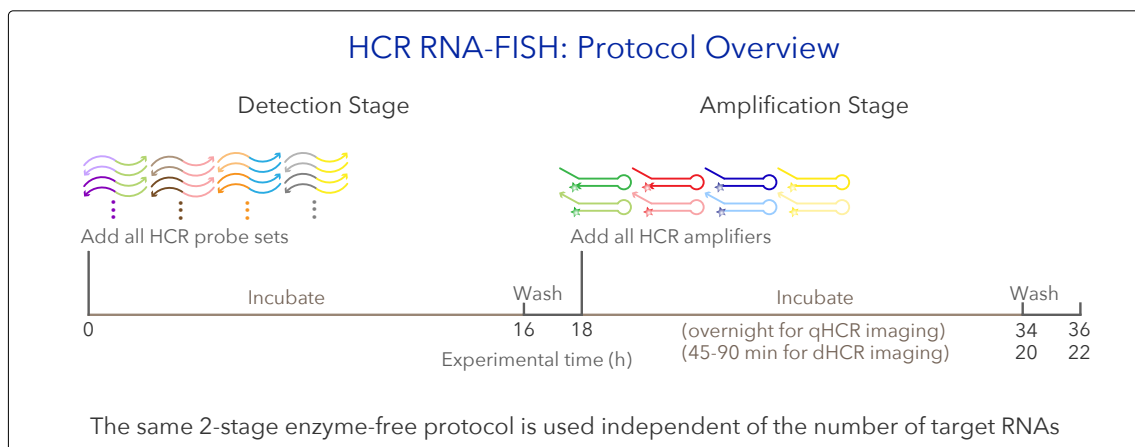
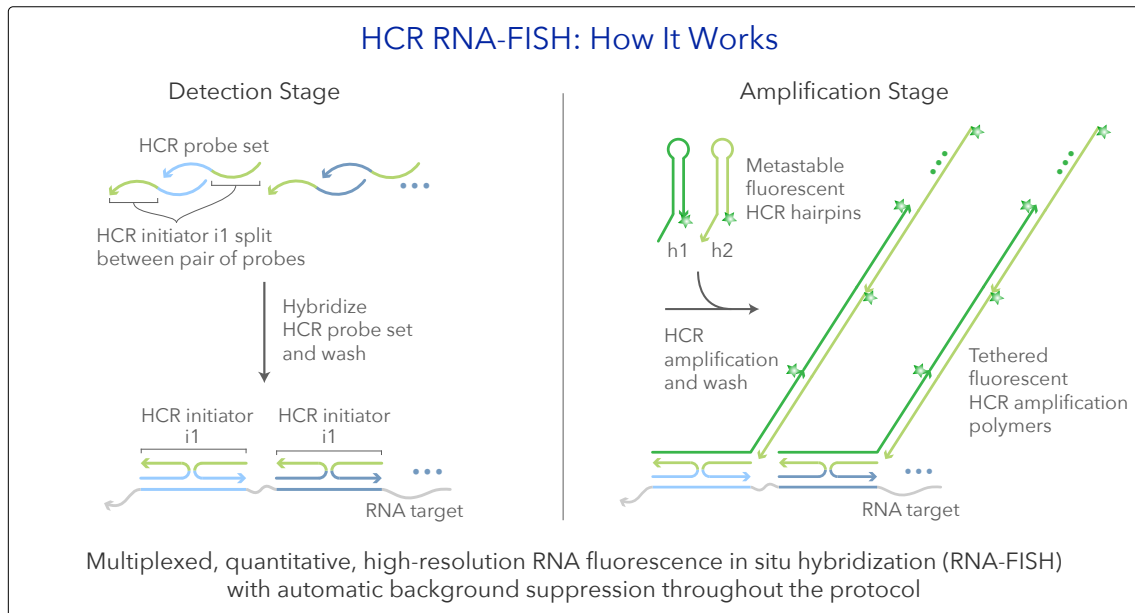
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## **HCR RNA-FISH**

*Multiplexed, quantitative, high-resolution RNA imaging*

### **Multiplexed Experiment**

- Order one HCR RNA-FISH bundle per target RNA

### **Example 2-Plex Experiment**

- HCR bundle for target mRNA1
  - HCR split-initiator probe set: target mRNA1 for use with amplifier B1
  - HCR amplifier: B1-647
  - HCR RNA-FISH buffers: probe hybridization buffer, probe wash buffer, amplification buffer (for use with all bundles)
- HCR bundle for target mRNA2
  - HCR split-initiator probe set: target mRNA2 for use with amplifier B2
  - HCR amplifier: B2-488

### **Storage conditions**

- Store HCR probe sets, HCR amplifiers, HCR probe hybridization buffer, and HCR probe wash buffer at -20 °C.
- Store HCR amplification buffer at 4 °C.
- On the bench top, keep stock solutions on ice.
- Make sure all solutions are well mixed before use.

## Preparation of fixed whole-mount sea urchin embryos

1. Incubate fertilized embryos with fresh filtered seawater in a 24-well plate at 16 °C until 45 hpf.
2. Transfer embryos from the 24-well plate into a 50 mL conical tube.
3. Centrifuge at  $100 \times g$  for 2 min to bring embryos to the bottom of the tube.
4. Remove seawater to reach 20 mL volume and add 2.4 mL of 4.45 M NaCl to deciliate embryos.
5. Incubate on ice until embryos settle to the bottom of the tube.
6. Remove seawater (with NaCl) and add 20 mL of filtered sea water.
7. Incubate on ice until embryos settle to the bottom of the tube.
8. Repeat steps 6 and 7.
9. Aspirate as much sea water as possible without removing embryos.
10. Fix embryos in 40 mL of 4% paraformaldehyde (PFA) overnight at 4 °C on a nutator.  
*CAUTION: use PFA with extreme care as it is a hazardous material.*  
*NOTE: use fresh PFA and cool to 4 °C before use to avoid increased autofluorescence.*
11. Remove fixative to reach a volume of  $\approx 6$  mL.
12. Transfer embryos to six 1.5 mL tubes (1 mL each).
13. Centrifuge at  $100 \times g$  for 2 min.
14. Wash embryos 5 times with 1 mL of PBST each. Centrifuge at  $100 \times g$  for 2 min to bring embryos to the bottom of a tube in between washes.
15. Wash embryos 3 times with 1 mL of 70% ethanol each. Centrifuge at  $100 \times g$  for 2 min to bring embryos to the bottom of a tube in between ethanol washes.
16. Store embryos in 1 mL of 70% ethanol at -20 °C before use.

## Multiplexed HCR RNA-FISH protocol

### Detection stage

1. Transfer  $\approx 200$  embryos to each well of a 96 well plate.

*NOTE: Adding and removing solution should be performed under a dissecting scope to avoid losing a large number of embryos.*

2. Rehydrate embryos  $3 \times 5$  min with 250  $\mu\text{L}$  of  $5\times$  SSCT.

3. Aspirate with care to reach  $\approx 10$   $\mu\text{L}$  of volume.

4. Add 50  $\mu\text{L}$  of probe hybridization buffer and pre-hybridize for 30 min at 37 °C.

*CAUTION: probe hybridization buffer contains formamide, a hazardous material.*

5. Prepare probe solution by adding 0.4 pmol of each probe set (e.g. 0.4  $\mu\text{L}$  of 1  $\mu\text{M}$  stock) to 50  $\mu\text{L}$  of probe hybridization buffer at 37 °C.

*NOTE: For dHCR imaging, use higher probe concentration (e.g., 16 nM) to increase probe hybridization yield. If desired, this approach can also be used to increase signal for qHCR imaging.*

6. Add the probe solution to reach a final hybridization volume of 100  $\mu\text{L}$ .

7. Gently stir the solution with a 10  $\mu\text{L}$  pipette tip.

8. Cover plate with Bio-Rad Microseal 'A' film and incubate embryos overnight (12–16 h) at 37 °C.

9. Add 150  $\mu\text{L}$  of probe wash buffer to each well of embryos.

*CAUTION: probe wash buffer contains formamide, a hazardous material.*

*NOTE: pre-heat probe wash buffer to 37 °C before use.*

10. Incubate at 37 °C for 5 min.

11. Remove excess probes by washing with  $\approx 200$   $\mu\text{L}$  of probe wash buffer at 37 °C:

(a)  $2 \times 5$  min

(b)  $2 \times 30$  min

*NOTE: Fill probe wash buffer to top of each well but do not overfill.*

*NOTE: It is important to maintain plate temperature at 37 °C during probe washing steps. This can be achieved by placing the 96-well plate containing the embryos on a heat plate while removing solution from each well.*

12. Wash embryos  $2 \times 5$  min with  $\approx 200$   $\mu\text{L}$  of  $5\times$  SSCT.

## Amplification stage

1. Separately prepare 6 pmol of hairpin h1 and 6 pmol of hairpin h2 by snap cooling 2  $\mu\text{L}$  of 3  $\mu\text{M}$  stock (heat at 95  $^{\circ}\text{C}$  for 90 seconds and cool to room temperature in a dark drawer for 30 min).  
*NOTE: HCR hairpins h1 and h2 are provided in hairpin storage buffer ready for snap cooling. h1 and h2 should be snap cooled in separate tubes.*
2. Equilibrate amplification buffer to room temperature.
3. Prepare hairpin solution by adding snap-cooled h1 hairpins and snap-cooled h2 hairpins to 100  $\mu\text{L}$  of amplification buffer at room temperature.
4. Aspirate as much 5 $\times$  SSCT as possible without removing embryos.
5. Add the hairpin solution and incubate embryos overnight (12–16 h) in the dark at room temperature.  
*NOTE: For dHCR imaging, amplify for a shorter period of time to ensure single-molecule dots are diffraction-limited.*
6. Add 150  $\mu\text{L}$  of 5 $\times$  SSCT and incubate for 5 min at room temperature.
7. Remove excess hairpins by washing with  $\approx 200$   $\mu\text{L}$  of 5 $\times$  SSCT at room temperature:
  - (a) 2  $\times$  5 min
  - (b) 2  $\times$  30 min
8. Samples can be stored at 4  $^{\circ}\text{C}$  protected from light before microscopy.

## Buffer recipes

### 4% Paraformaldehyde (PFA)

4% PFA  
1× PBS

### For 40 mL of solution

10 mL of 16% PFA solution  
4 mL of 10× PBS  
Fill up to 40 mL with ultrapure H<sub>2</sub>O

### PBST

1× PBS  
0.1% Tween 20

### For 50 mL of solution

5 mL of 10× PBS  
500 μL of 10% Tween 20  
Fill up to 50 mL with ultrapure H<sub>2</sub>O

### 5× SSCT

5× sodium chloride sodium citrate (SSC)  
0.1% Tween 20

### For 40 mL of solution

10 mL of 20× SSC  
400 μL of 10% Tween 20  
Fill up to 40 mL with ultrapure H<sub>2</sub>O

*NOTE: 16% PFA solution is filter sterilized using a 25 mm syringe filter with 0.2 μm membrane before use.*

*NOTE: avoid using calcium chloride and magnesium chloride in PBS as this leads to increased autofluorescence in the embryos.*

## HCR Technology Citation Notes

For citation, please select from the list below as appropriate for your application:

- **HCR IHC + HCR RNA-FISH**

[HCR IHC + HCR RNA-FISH](#) enables a unified approach to multiplexed, quantitative, high-resolution protein immunohistochemistry (IHC) and RNA fluorescence in situ hybridization (RNA-FISH), with quantitative 1-step enzyme-free HCR signal amplification performed for all protein and RNA targets simultaneously ([Schwarzkopf et al., 2021](#)).

- **HCR IHC**

[HCR IHC](#) enables multiplexed, quantitative, high-resolution protein immunohistochemistry (IHC) in highly autofluorescent samples (e.g., FFPE brain tissue sections) ([Schwarzkopf et al., 2021](#)).

- **HCR RNA-FISH (v3.0)**

Third-generation [HCR RNA-FISH \(v3.0\)](#) enables multiplexed, quantitative, high-resolution RNA fluorescence in situ hybridization (RNA-FISH) with [automatic background suppression throughout the protocol](#) for dramatically enhanced performance (signal-to-background, qHCR precision, dHCR fidelity) and ease-of-use (no probe set optimization for new targets and organisms) ([Choi et al., 2018](#)). Quantitative analysis modes:

- [qHCR RNA imaging](#): analog mRNA relative quantitation with subcellular resolution in the anatomical context of thick autofluorescent samples.
- [dHCR RNA imaging](#): digital mRNA absolute quantitation with single-molecule resolution in the anatomical context of thick autofluorescent samples.
- [qHCR RNA flow cytometry](#): analog mRNA relative quantitation for high-throughput expression profiling of mammalian cells and bacteria.

[Protocols for HCR RNA-FISH \(v3.0\)](#) in diverse organisms are adapted from the Zoo paper.

- **qHCR RNA imaging**

[qHCR RNA imaging](#) enables mRNA relative quantitation with subcellular resolution in the anatomical context of thick autofluorescent samples (e.g., whole-mount vertebrate embryos). The [read-out/read-in analysis framework](#) enables bidirectional quantitative discovery in an anatomical context ([Trivedi et al., 2018](#)).

- **Zoo paper**

Protocols for multiplexed mRNA imaging in diverse sample types ([Choi et al., 2016](#)):

- bacteria in suspension
- FFPE human tissue sections
- generic sample in solution
- generic sample on a slide
- mammalian cells on a slide
- mammalian cells in suspension
- whole-mount chicken embryos
- whole-mount fruit fly embryos
- whole-mount mouse embryos
- whole-mount nematode larvae
- whole-mount sea urchin embryos
- whole-mount zebrafish embryos and larvae

- **dHCR imaging**

[dHCR RNA imaging](#) enables RNA absolute quantitation with single-molecule resolution in the anatomical context of thick autofluorescent samples (e.g., 0.5 mm adult mouse brain sections) ([Shah et al., 2016](#)).

- **qHCR northern blots**

[qHCR northern blots](#) enable multiplexed quantification of RNA target size and abundance for up to 5 target RNAs ([Schwarzkopf & Pierce, 2016](#)).

- **HCR RNA-FISH (v2.0)**

Second-generation in situ HCR RNA-FISH technology (v2.0) using DNA HCR probes and DNA HCR probes and DNA HCR amplifiers: 10× increase in signal, 10× reduction in cost, dramatic increase in reagent durability ([Choi et al., 2014](#)).

- **HCR RNA-FISH (v1.0)**

First-generation HCR RNA-FISH technology (v1.0) using RNA HCR probes and RNA HCR amplifiers: multiplexed mRNA imaging in whole-mount vertebrate embryos with simultaneous signal amplification for up to 5 target mRNAs ([Choi et al., 2010](#)).

- **HCR mechanism**

The [hybridization chain reaction \(HCR\) mechanism](#) enables multiplexed, quantitative, 1-step, isothermal, enzyme-free signal amplification in diverse technological settings ([Dirks & Pierce, 2004](#)).



## HCR Technology References

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